



MERCURIUS™

**Organoid DRUG-seq Service
(Frozen organoid pellet)**

**Sample Preparation and
Submission Guidelines**

RNA Extraction-free Protocol for
96- and 384-well Plate Format

January 2025

Sample Submission Guidelines at a glance

1. Wash the organoids and store them in the original culturing plates at -80°C before shipment.
2. Fill in the Sample Submission Form (**SSF**) and **check all the boxes** in the Sample Submission checklist below; send both files to **orders@alitheagenomics.com**. Please be aware that any inconsistency may lead to delays or additional fees.
3. Ship the samples on dry ice and send us the shipment tracking number.

Sample submission checklist

- The Sample Submission Form (SSF) must be filled properly with a unique sample ID. Consider adding a suffix for technical replicates (e.g., XX_rep1, XX_rep2, etc.). Ensure that SSF provides information about all shipped samples.
- The **minimum number** of samples in each group (to be pooled together) is **16** (for 96-well plate) and **150** (for 384-well plate).
- One type of seeded cell per well and only one type to be pooled together, with a minimum of 80'000 cells per pool.
- Sample plates are labeled with the same Plate ID as in the Sample Submission Form.
- Plates are well sealed with the Aluminum or plastic seal.

Consumables not provided

Reagents	Manufacturer	PN
DPBS, no calcium, no magnesium	Gibco	14190144
Nuclease-free water	Thermo	A57775

1. Essential considerations for input material

- 1.1 The organoids lysate protocol was validated with organoids and spheroids, cultured in **flat-bottom 96-well plates with Matrigel®** or **microtissue-specific 96-well plates** (e.g., Akura™, BIOFLOAT™, ULA Corning plates, etc.)
- 1.2. The recommended input range of cells in organoids or spheroids is 5'000-50'000 cells/well of 96WP and 2'000-10'000 cells/well of 384WP.
- 1.3. Organoids must be plated a few days before for the best results.
- 1.4. Depending on the type of organoids or spheroids (human, mouse, metastatic, or primary cells) and experimental design (e.g., drug treatment, induction of apoptosis, cell cycle arrest, etc.), consider the organoid-specific cell doubling time and the potential effect of the treatment on cell quality and quantity.

2. Organoid Pellets Preparation for input material

- 2.1 Gently aspirate the culture media from the plate and wash organoids or spheroids by adding DPBS (per well):
 - 100 µL for **96WP flat-bottom plates with Matrigel®**,
 - 20 µL for **96WP microtissue-specific plates and 384WP**.
- 2.2 In case of spheroids, centrifuge plate at 300x g for 5 min.
- 2.3 Gently tap the plate and aspirate as much DPBS as possible without disturbing the structure of the organoids.
- 2.4 Seal the plate well with an Aluseal and immediately transfer it to a -80°C freezer for storage. If possible, snap-freeze the plate with dry ice beforehand.
- 2.5 Store the plate at -80°C before shipment.

NOTE: If several plates must be processed, perform the procedure individually per plate to avoid keeping the plates at room temperature for a prolonged time.

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